



Shipley ON, Polunin NVC, Newman SP, Sweeting CJ, Barker S, Witt MJ, Brooks EJ.

[Stable isotopes reveal insight into food web dynamics of a data-poor deep-sea island slope community.](#)

*Food Webs 2017,*

<http://dx.doi.org/10.1016/j.fooweb.2017.02.004>

**Copyright:**

© 2017. This manuscript version is made available under the [CC-BY-NC-ND 4.0 license](#)

**DOI link to article:**

<http://dx.doi.org/10.1016/j.fooweb.2017.02.004>

**Date deposited:**

14/02/2017

**Embargo release date:**

12 February 2018



This work is licensed under a

[Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International licence](#)

## Accepted Manuscript

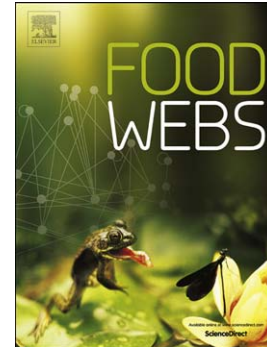
Stable isotopes reveal insight into food web dynamics of a data-poor deep-sea island slope community

Oliver N. Shipley, Nicholas V.C. Polunin, Steven P. Newman, Christopher J. Sweeting, Sam Barker, Matthew J. Witt, Edward J. Brooks

PII: S2352-2496(16)30029-5  
DOI: doi:[10.1016/j.fooweb.2017.02.004](https://doi.org/10.1016/j.fooweb.2017.02.004)  
Reference: FOOWEB 50

To appear in:

Received date: 6 December 2016  
Revised date: 20 January 2017  
Accepted date: 3 February 2017



Please cite this article as: Shipley, Oliver N., Polunin, Nicholas V.C., Newman, Steven P., Sweeting, Christopher J., Barker, Sam, Witt, Matthew J., Brooks, Edward J., Stable isotopes reveal insight into food web dynamics of a data-poor deep-sea island slope community, (2017), doi:[10.1016/j.fooweb.2017.02.004](https://doi.org/10.1016/j.fooweb.2017.02.004)

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

# Stable isotopes reveal insight into food web dynamics of a data-poor deep-sea island slope community

Oliver N. Shipley<sup>1, 2, 3\*</sup>, Nicholas V. C. Polunin<sup>1</sup>, Steven P. Newman<sup>1, 4</sup>, Christopher J. Sweeting<sup>1</sup>, Sam Barker<sup>5</sup>, Matthew J. Witt<sup>5</sup> and Edward J. Brooks<sup>2</sup>

<sup>1</sup>School of Marine Science & Technology, Newcastle University, Newcastle upon Tyne NE1 7RU UK

<sup>2</sup>Shark Research and Conservation Program, Cape Eleuthera Institute, Eleuthera, The Bahamas, PO Box EL-26029

<sup>3</sup>School of Marine and Atmosphere Sciences, Stony Brook University, 11780 NY, USA

<sup>4</sup>Banyan Tree Marine Lab, Vabbinfaru, Kaafu Atoll, Maldives

<sup>5</sup>The Environment and Sustainability Institute, University of Exeter Penryn Campus, Cornwall, TR10 9EZ, UK

\*Corresponding author

## Abstract

Deep-sea communities are subject to a growing number of extrinsic pressures, which threatens their structure and function. Here we use carbon and nitrogen stable isotopes to provide new insights into the community structure of a data-poor deep-sea island slope system, the Exuma Sound, The Bahamas. A total of 78 individuals from 16 species were captured between 462 m and 923 m, and exhibited a broad range of  $\delta^{13}\text{C}$  (9.45‰) and  $\delta^{15}\text{N}$  (6.94‰). At the individual-level,  $\delta^{13}\text{C}$  decreased strongly with depth, indicative of shifting production sources, as well as potential shifts in community composition, and species-specific feeding strategies.  $\delta^{15}\text{N}$  did not follow strong depth relationships, suggesting trophic level and depth are not tightly coupled across individuals. We observed ontogenetic enrichment in  $\delta^{13}\text{C}$  and  $\delta^{15}\text{N}$  for *Squalus cubensis* (Cuban dogfish) highlighting a shift to larger, higher trophic level prey through ontogeny. These data provide the first assessment of food-web structure in the Exuma Sound, and suggest inherent complexity associated with deep-sea island slope ecosystems. Such observations are needed to further our understanding and develop contemporary management plans for these systems.

**Keywords:** Stable isotope analysis; deep-water; ecosystem; primary production; feeding dynamics

## 1. Introduction

Deep-sea ecosystems are being increasingly exploited by a multitude of anthropogenic activities, such as commercial fishing and mining, which threatens their structure and function (Benn et al., 2010; Priede et al., 2010; Van Dover et al., 2011). The conservative life histories of many deep-sea taxa exacerbate exploitative effects, as many are unable to quickly rebound from population-based perturbations (Simpfendorfer and Kyne, 2009). Such effects are likely to drive changes in community composition, and structure, as seen in a number of coastal biomes (e.g. McCain et al., 2015; Möllman et al., 2015). Therefore a basic understanding of food-web dynamics is essential for predicting the broad-scale impacts of exploitation on deep-sea communities.

Stable isotope analysis (SIA) of carbon and nitrogen (referred to in delta notation as  $\delta^{13}\text{C}$  and  $\delta^{15}\text{N}$ ) has proven valuable in describing food-web structure and species interactions across marine, freshwater, and terrestrial biomes (Layman et al., 2011; Hussey et al., 2012; Rundel et al., 2012), and are becoming increasingly applied to deep-sea systems (Polunin et al., 2001; Pethybridge et al., 2012; Churchill et al., 2015; Shipley et al., In Press). Although the majority of isotopic food-web studies have focussed on shallow water systems, evidence suggests deep-sea

community structure is highly complex (Portail et al., 2016), can encompass many trophic guilds (Valls et al., 2014), and is often supported by multiple sources of primary production (Newman et al., 2011; Reid et al., 2012). Deep-sea island slope communities are of particular interest to managers, as they occur at the interface between productive neritic and more oligotrophic open-ocean systems. Given their proximity to a multitude of potential energy pools, these systems may exhibit a unique suite of community dynamics and support a highly diverse range of taxa (Suchanek et al., 1985). Despite this, deep-sea island slopes are amongst the most poorly represented across the scientific literature (Shipley et al., *In Press*), highlighting a critical need to further examine these systems.

The Exuma Sound is a deep-water (~1700m max depth) inlet of the Atlantic Ocean, categorised by narrow continental shelves, which drop from 30m to 400m at the shelf-break. The continental slope is categorised by rugose limestone outcroppings creating a complex bathymetry (Ball et al., 1969), which transitions into a featureless sandy benthos at the bottom (Crevello and Schlager, 1980). Data are sparse, but the system appears to support high megafaunal diversity and abundance (Brooks et al., 2015). Here we use carbon and nitrogen stable isotopes to explore the food-web structure of the Exuma Sound deep-sea island slope community. We provide the first reported  $\delta^{13}\text{C}$  and  $\delta^{15}\text{N}$  values for a number of species, and explore potential factors responsible for patterns in isotope values across individuals.

## 2. Methods

Sampling was conducted in the northeast Exuma Sound, off south Eleuthera, The Bahamas (24.54°N, 76.12°W; WGS84), between November 2013 and May 2014, under permits MAF/FIS/17 and MAF/FIS/34 from the Bahamian Department of Marine Resources. Animal sampling followed the requirements of the UK Home Office Scientific Procedures (Animals) Act and Association for the Study of Animal Behaviour (Rollin and Kessel, 1998). Animals were captured using deep-sea longlines (see Brooks et al., 2015 for detailed methods) and square and cylindrical baited fish traps soaked for 24 hours. White muscle tissue, which provides a relatively long-term (weeks to months) indication of diet (Hussey et al., 2012) was excised dorsally for fish and eels, and taken from appendages for invertebrates. Tissue was then stored at -20°C prior to SIA. Tissues were freeze-dried and homogenised prior to polar compound (lipid, urea and trimethylamine N-oxide (TMAO)) extraction with a 2:1 chloroform methanol solution (Sweeting et al., 2006; Churchill et al., 2015). Approximately  $0.6 \pm 0.1$  mg of dried tissue was then weighed into 6x4 mm tin-capsules.

All samples were analysed in duplicate for  $\delta^{15}\text{N}$ ,  $\delta^{13}\text{C}$ , %N and %C using a Sercon INTEGRA2 stable isotope mass spectrometer (SerCon Ltd, Cheshire, UK) at the Environment and Sustainability Institute, University of Exeter, Penryn Campus, UK. Values are referred to in  $\delta$  notation following the equation:  $\delta^b\text{X}[(R_{\text{sample}}/R_{\text{standard}})-1] \times 1000$  where  $X^b$  = isotopic element chosen for analysis,  $R_{\text{sample}}$  = heavy to light isotope ratio derived from the sample, and  $R_{\text{standard}}$  = heavy to light isotope ratio derived from the standard (DeNiro and Epstein, 1978). Isotope ratio scales were normalised through Vienna Pee Dee Belemnite for carbon, and atmospheric  $\text{N}_2$  for nitrogen using IAEA standards of lithium carbonate (LSVEC), CH-6 and N-1, N-2 respectively. To determine analytical precision (standard deviation), two alanine standards were placed between every 8 samples ( $n = 162$ ,  $< 0.15$  S.D.). Batch precision was determined with an internal reference of blue antimora (*Antimora rostrata*) white muscle tissue, which was placed at the beginning and end of every run ( $n = 32$ ,  $< 0.15$  S.D.). A two-point linear correction (Paul et al., 2010) was applied to  $\delta^{15}\text{N}$  values based on IAEA N-1 and N-2 values. The accuracy of the carbon IAEA reference materials was such that a two point linear correction was not necessary.

At the individual level, Generalized Linear Models (GLM's) were used to determine whether variation in  $\delta^{13}\text{C}$  and  $\delta^{15}\text{N}$  was best explained by depth, species, or an interaction between both variables. For *S. cubensis*, body-size relationships with  $\delta^{13}\text{C}$  and  $\delta^{15}\text{N}$  were assessed using least-squares regression. All statistical analyses were performed in 'R' (Version 3.0; R development core team, 2007).

### 3. Results

A total of 78 individuals from 16 species were captured from the northeast Exuma Sound, at depths ranging from 462 m to 923 m (Table 1). At the individual level, community  $\delta^{13}\text{C}$ ,  $\delta^{15}\text{N}$  and C:N ranged from -10.77‰ to -20.22‰, 6.99-13.93‰, and 2.57 to 3.34 respectively (Table 1).

The effect of depth (glm,  $t = -4.62$ ,  $p < 0.001$ ), species ( $t = -2.48$ ,  $p < 0.02$ ), and interaction of the two factors ( $t = 2.57$ ,  $p < 0.02$ ) all proved significant in explaining variation in  $\delta^{13}\text{C}$ , which became more depleted in individuals at greater depths. Variation in  $\delta^{15}\text{N}$  could not be explained by depth ( $t = 0.41$ ,  $p > 0.15$ ), species ( $t = 1.02$ ,  $p > 0.15$ ), or interaction of both factors ( $t = 0.81$ ,  $p > 0.15$ ). For *Squalus cubensis*  $\delta^{13}\text{C}$  (least-squares regression,  $r^2 = 0.69$ ,  $p < 0.001$ ) and  $\delta^{15}\text{N}$  (least-squares regression,  $r^2 = 0.26$ ,  $p < 0.001$ ) both became enriched with increasing size.

**Table 1.**

**Fig 1.**

### 4. Discussion

The wide range of  $\delta^{13}\text{C}$  (9.45‰) and  $\delta^{15}\text{N}$  (6.94‰) values across individuals suggests a unique and complex food web structure, spanning a number of trophic levels in which biomass may be underpinned by a mix of potential production sources. At the individual level,  $\delta^{13}\text{C}$  became more depleted in individuals captured at greater depths. Such patterns could be explained by a number of factors including shifting production sources, or shifts in community composition and thus faunal-specific feeding strategies (i.e. switching from benthic to pelagic prey) (Trueman et al., 2014). The major energy pools supporting deep-sea food-web productivity are poorly defined, aside from oligotrophic open-ocean (phytoplanktonic-POM) and hydrothermal vent (Chemoautotrophic and phytoplanktonic POM) communities. However, a significant proportion of neritic-derived primary production may be transported into deep-sea systems at some locales (Suchanek et al., 1985; Sanchez-Vidal et al., 2012), and if assimilated into the food web, could drive the more enriched  $\delta^{13}\text{C}$  values found in shallower individuals.

**Fig 2.**

As the variability in  $\delta^{13}\text{C}$ , was also part explained by species, and the interaction between species and depth, a shift in community composition, could also part-explain the depletion in  $\delta^{13}\text{C}$  with depth, especially for fishes. *M. canis-insularis*, which was captured shallowest, is known to feed largely on benthic crustaceans (Gelsleichter et al., 1999), while *Merluccius albidus*, *Hexanchus nakamurai*, *S. cubensis*, and *Centrophorus* spp., which are found deeper, feed on more pelagic prey (teleosts and squid) (Compagno, 1984; Cohen et al., 1990; Monzini, 2006; Ebert et al., 2009). Furthermore, some deep-sea fishes are known to perform considerable vertical movements throughout the water column (e.g. Shipley et al., 2017), which could facilitate benthic-pelagic coupling, thus transporting more isotopically depleted resources from the pelagic zone to greater depths on the slope (Trueman et al., 2014). The size-based enrichment of  $\delta^{13}\text{C}$  and  $\delta^{15}\text{N}$  exhibited by *S. cubensis* suggests prey switching throughout ontogeny, as larger individuals are able to ingest larger, and a more diverse suite of prey items, as in the Gulf of Mexico (Churchill et al., 2015).

Isotope data from the Exuma Sound also provide insights into the feeding dynamics of invertebrates. The high  $\delta^{15}\text{N}$  values in isopods, for example *Bathynomus giganteus*, agree with observations from the Gulf of Mexico, and may attribute to a greater degree of scavenging compared to *Heterocarpus ensifer* (McAvoy et al., 2002; Churchill, 2015). High  $\delta^{15}\text{N}$  in isopods may also be attributable to their broad depth range. This suggests individuals scavenge over broader regions of the slope, which increases the chance of encountering prey items exhibiting more enriched  $\delta^{15}\text{N}$ . Isopods can often be micropredators of larger fishes (Shipley et al., 2016), which would further explain high  $\delta^{15}\text{N}$  values.

The large ranges, and depth patterns of both  $\delta^{13}\text{C}$  and  $\delta^{15}\text{N}$  indicate complexity within the Exuma Sound community. We also highlight how multiple factors, such as a changing mix of production sources, and changing community composition may directly affect  $\delta^{13}\text{C}$  of deep-sea fauna. Future work should therefore focus on defining energy specific energy sources underpinning deep-sea communities, as well as examining clear predator-prey relationships across species. Finally, examining similar systems at other locales is suggested if the ecological significance and energy sources supporting these communities are to be better understood.

## 5. Acknowledgements

Thanks to the Cape Eleuthera Foundation for funding fieldwork. We also thank B. Talwar, M. Violich, O. O'Shea, I. Bouyoucos, L. Charash, K. Church, C. Mounts, J. McAlevey, C. Valente and J. Forster for field assistance. D. Whittaker and P. MacParlane provided laboratory assistance, and the University of Exeter funded the stable isotope analysis.

## 6. Literature Cited

- Ball, M. M., Harrison, C. G. A., Hurley, R. J., & Leist, C. E. (1969). Bathymetry in the vicinity of the northeastern scarp of the Great Bahama Bank and Exuma Sound. *Bulletin of Marine Science* 19:243-252.
- Benn, A. R., Weaver, P. P., Billet, D. S., Van Den Hove, S., Murdock, A. P., Doneghan, G. B., & Le Bas, T. (2010). Human activities on the deep seafloor in the North East Atlantic: an assessment of spatial extent. *PLoS ONE* 5:e12730.
- Brooks, E. J., Brooks, A. M., Williams, S., Jordan, L. K., Abercrombie, D., Chapman, D. D., Howey-Jordan, L. A., & Grubbs, R. D. 2015. First description of deep-water elasmobranch assemblages in the Exuma Sound, The Bahamas. *Deep Sea Research Part II: Topical Studies in Oceanography* 115:81-91.
- Buchan, K. C. (2000). The Bahamas. *Marine Pollution Bulletin* 41: 94-111.
- Compagno, L.J.V., 1984. *FAO Species Catalogue*. Vol. 4. *Sharks of the world. An annotated and illustrated catalogue of shark species known to date. Part 1 - Hexanchiformes to Lamniformes*. *FAO Fisheries Synopses*. 125:1-249.
- Churchill, D. A. (2015) *Investigating Trophic Interactions of Deep-sea Animals (Sharks, Teleosts, and Mobile scavengers) in the Gulf of Mexico Using Stable Isotope Analysis*. FIU Electronic Theses and Dissertations. Paper 2214.
- Churchill, D. A., Heithaus, M. R., Vaudo, J. J., Grubbs, R. D., Gastrich, K., & Castro, J. I. (2015). Trophic interactions of common elasmobranchs in deep-sea communities of the Gulf of Mexico revealed through stable isotope and stomach content analysis. *Deep Sea Research. Part II: Topical Studies in Oceanography* 115:92-102.
- Cohen, D.M., T. Inada, T. Iwamoto and N. Scialabba, 1990. *FAO species catalogue*. Vol. 10. *Gadiform fishes of the world (Order Gadiformes)*. An annotated and illustrated catalogue of

- cods, hakes, grenadiers and other gadiform fishes known to date. FAO Fisheries Synopses. 125(10).
- Crevello, P. D., & Schlager, W. (1980). Carbonate debris sheets and turbidites, Exuma Sound, Bahamas. *Journal of Sedimentary Research* 50: 1121-1147.
- DeNiro, M. J., & Epstein, S. 1978. Influence of diet on the distribution of carbon isotopes in animals. *Geochimica et cosmochimica acta* 42:495-506.
- DeNiro, M. J., & Epstein, S. (1981). Influence of diet on the distribution of nitrogen isotopes in animals. *Geochimica et cosmochimica acta* 45:341-351.
- Ebert, D.A., Serena, F. & Mancusi, C. 2009. *Hexanchus nakamurai*. The IUCN Red List of Threatened Species 2009 (<http://dx.doi.org/10.2305/IUCN.UK.2009-2.RLTS.T161352A5404404.en>)
- Gelsleichter, J., Musick, J. A., & Nichols, S. (1999). Food habits of the smooth dogfish, *Mustelus canis*, dusky shark, *Carcharhinus obscurus*, Atlantic sharpnose shark, *Rhizoprionodon terraenovae*, and the sand tiger, *Carcharias taurus*, from the northwest Atlantic Ocean. *Environmental Biology of Fishes* 54:205-217.
- Hussey, N. E., MacNeil, M. A., Olin, J. A., McMeans, B. C., Kinney, M. J., Chapman, D. D., & Fisk, A. T. (2012). Stable isotopes and elasmobranchs: tissue types, methods, applications and assumptions. *Journal of Fish Biology* 80:1449-1484.
- Layman, C. A., Araujo, M. S., Boucek, R., Hammerschlag- Peyer, C. M., Harrison, E., Jud, Z. R., ... & Post, D. M. (2012). Applying stable isotopes to examine food- web structure: an overview of analytical tools. *Biological Reviews*: 87:545-562.
- MacAvoy, S. E., Carney, R. S., Fisher, C. R., & Macko, S. A. (2002). Use of chemosynthetic biomass by large, mobile, benthic predators in the Gulf of Mexico. *Marine Ecology Progress Series* 225:65-78.
- McCain, J. S. P., Cull, D. J., Schneider, D. C., & Lotze, H. K. (2015). Long-term shift in coastal fish communities before and after the collapse of Atlantic cod (*Gadus morhua*). *ICES Journal of Marine Science*:fsv126
- McCauley, D. J., Young, H. S., Dunbar, R. B., Estes, J. A., Semmens, B. X., & Micheli, F. (2012). Assessing the effects of large mobile predators on ecosystem connectivity. *Ecological Applications* 22:1711-1717.
- Möllmann, C., Folke, C., Edwards, M., & Conversi, A. (2015). Marine regime shifts around the globe: theory, drivers and impacts. *Philosophical Transactions of the Royal Society B: Biological Sciences* 370:1659.
- Monzini, J. 2006. *Squalus cubensis*. The IUCN Red List of Threatened Species 2006: (<http://dx.doi.org/10.2305/IUCN.UK.2006.RLTS.T61416A12476876.en>)
- Myers, R. A., Baum, J. K., Shepherd, T. D., Powers, S. P., & Peterson, C. H. (2007). Cascading effects of the loss of apex predatory sharks from a coastal ocean. *Science* 315:1846-1850.
- Newman, M. C., Xu, X., Cotton, C. F., & Tom, K. R. (2011). High mercury concentrations reflect trophic ecology of three deep-water chondrichthyans. *Archives of Environmental Contamination and Toxicology* 60:618-625.
- Papastamatiou, Y., Meyer, C. G., Kosaki, R. K., Wallsgrove, N. J., & Popp, B. N. (2015). Movements and foraging of predators associated with mesophotic coral reefs and their potential for linking ecological habitats. *Marine Ecology Progress Series* 52:155 – 170.
- Paul, D., Skrzypek, G., & Forizs, I. 2007. Normalization of measured stable isotopic compositions to isotope reference scales—a review. *Rapid Communications in Mass Spectrometry* 21:3006-3014.

- Pethybridge, H., Butler, E. C. V., Cossa, D., Daley, R., & Boudou, A. (2012). Trophic structure and biomagnification of mercury in an assemblage of deepwater chondrichthyans from southeastern Australia. *Marine Ecology Progress Series* 451:163-174.
- Pinnegar, J. K., & Polunin, N. V. C. (1999). Differential fractionation of  $\delta^{13}\text{C}$  and  $\delta^{15}\text{N}$  among fish tissues: implications for the study of trophic interactions. *Functional Ecology* 13:225-231.
- Polunin, N. V. C., Morales-Nin, B., Pawsey, W. E., Cartes, J. E., Pinnegar, J. K., & Moranta, J. (2001). Feeding relationships in Mediterranean bathyal assemblages elucidated by stable nitrogen and carbon isotope data. *Marine Ecology Progress Series* 220:13-23.
- Portail, M., Olu, K., Dubois, S. F., Escobar-Briones, E., Gelinas, Y., Menot, L., & Sarrazin, J. (2016). Food-Web Complexity in Guaymas Basin Hydrothermal Vents and Cold Seeps. *Plos One* 11: e0162263.
- Priede, I. G., Godbold, J. A., Niedzielski, T., Collins, M. A., Bailey, D. M., Gordon, J. D., & Zuur, A. F. (2010). A review of the spatial extent of fishery effects and species vulnerability of the deep-sea demersal fish assemblage of the Porcupine Seabight, Northeast Atlantic Ocean. *ICES Journal of Marine Science: Journal du Conseil* 68:281-289
- R Development Core Team. 2007. R: a language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria.
- Rankey, E. C., & Reeder, S. L. (2011). Holocene oolitic marine sand complexes of the Bahamas. *Journal of Sedimentary Research* 81:97-117.
- Reid, W. D., Wigham, B. D., McGill, R. A., & Polunin, N. V.C. (2012). Elucidating trophic pathways in benthic deep-sea assemblages of the Mid-Atlantic Ridge north and south of the Charlie-Gibbs Fracture Zone. *Marine Ecology Progress Series* 463:89-103.
- Rollin, B. E., & Kessel, M. L. (1998). Guidelines for the treatment of animals in behavioural research and teaching. *Animal Behaviour* 55:251-257.
- Rundel, P., Ehleringer, J. R., & Nagy, K. A. (Eds.). (2012). *Stable isotopes in ecological research* (Vol. 68). Springer Science & Business Media.
- Sanchez-Vidal, A., Canals, M., Calafat, A. M., Lastras, G., Pedrosa-Pàmies, R., Menéndez, M., ... & Alcoverro, T. (2012). Impacts on the deep-sea ecosystem by a severe coastal storm. *PLoS One* 7:e30395.
- Shipley, O., Talwar, B., Grubbs, D., & Brooks, E. (2016). Isopods present on deep-water sharks *Squalus cubensis* and *Heptranchias perlo* from The Bahamas. *Marine Biodiversity*:1-2.
- Shipley, O. N., Howey, L. A., Tolentino, E. R., Jordan, L. K. B., & Brooks, E. J. (2017). Novel techniques and insights into the deployment of pop-up satellite archival tags on a small-bodied deep-water chondrichthyan. *Deep-Sea Research Part I: Oceanographic Research Papers* 119: 81 – 90.
- Shipley, O. N., Brooks, E. J., Madigan, D. J., Sweeting, C. J., Grubbs, R. D. (In Press). Stable isotope analysis in deep-sea chondrichthyans: recent challenges, ecological insights, and future directions. *Reviews in Fish Biology and Fisheries*.
- Simpfendorfer, C. A., & Kyne, P. M. (2009). Limited potential to recover from overfishing raises concerns for deep-sea sharks, rays and chimaeras. *Environmental Conservation* 36: 97-103.
- Suchanek, T. H., Williams, S. L., Ogden, J. C., Hubbard, D. K., & Gill, I. P. (1985). Utilization of shallow-water seagrass detritus by Caribbean deep-sea macrofauna:  $\delta^{13}\text{C}$  evidence. *Deep Sea Research Part A: Oceanographic Research Papers* 32:201-214.



- Sweeting, C. J., Polunin, N. V. C., & Jennings, S. (2006). Effects of chemical lipid extraction and arithmetic lipid correction on stable isotope ratios of fish tissues. *Rapid Communications in Mass Spectrometry* 20:595-601.
- Trueman, C. N., Johnston, G., O'Hea, B., & MacKenzie, K. M. (2014). Trophic interactions of fish communities at midwater depths enhance long-term carbon storage and benthic production on continental slopes. *Proceedings of the Royal Society of London B: Biological Sciences* 281: 20140669.
- Valls, M., Sweeting, C. J., Olivar, M. P., de Puellas, M. F., Pasqual, C., Polunin, N. V. C., & Quetglas, A. (2014). Structure and dynamics of food webs in the water column on shelf and slope grounds of the western Mediterranean. *Journal of Marine Systems*, 138:171-181.
- Van Dover, C. L. (2011). Tighten regulations on deep-sea mining. *Nature* 470: 31-33.

## 7. Tables and Figures

**Table 1** – Species, number of samples (*n*), mean  $\delta^{15}\text{N}$ ,  $\delta^{13}\text{C}$  and C:N ( $\pm$  SD) gathered from the Exuma Sound deep-sea slope community.

Species	Common Name	<i>n</i>	Capture Depth (m)	$\delta^{15}\text{N}$ ( $\pm$ SD)	$\delta^{13}\text{C}$ ( $\pm$ SD)	C:N ( $\pm$ SD)
<b>Elasmobranchs</b>						
<i>Squalus cubensis</i>	Cuban dogfish	22	462 - 727	9.98 (0.71)	-14.08 (0.70)	2.82 (0.08)
<i>Centrophorus</i> spp.	Gulper shark	9	731 – 818	10.62 (0.24)	-16.09 (0.24)	2.77 (0.12)
<i>Mustelus canis-insularis</i>	Smooth dogfish	6	462 – 655	9.10 (0.32)	-11.83 (0.91)	2.82 (0.10)
<i>Hexanchus nakamurai</i>	Bigeye sixgill shark	4	534 – 641	11.23 (0.05)	-13.80 (0.60)	2.89 (0.28)
<i>Heptranchias perlo</i>	Sharpnose sevengill shark	2	607 – 656	12.77 (0.56)	-16.49 (0.51)	2.86 (0.10)
<i>Scyliorhinus meadi</i>	Blotched catshark	1	602	8.03	-14.05	2.76
<b>Other Fish and Eels</b>						
<i>Synaphobranchus affinis</i>	Grey cutthroat eel	2	923 – 978	11.24 (0.59)	-15.42 (0.39)	3.15 (0.03)
<i>Sebastes oculatus</i>	Patagonian redfish	1	559	11.87	-14.42	2.86
<i>Merluccius albidus</i>	Offshore hake	1	731	13.93	-15.69	2.96
<i>Polymixia nobilis</i>	Stout beardfish	1	606	11.6	-13.6	2.94
<i>Rhomboplites aurorubens</i>	Vermillion snapper	1	461	10.07	-13.15	2.97
<i>Conger</i> sp.	Conger eel	1	748	12.05	-13.93	2.89
<i>Nemichthys scolopaceus</i>	Slender snipe eel	1	-	8.95	-16.19	2.97
<b>Invertebrates</b>						
<i>Bathynomus giganteus</i>	Giant isopod	10	656 - 912	12.57 (0.59)	-15.04 (0.51)	3.06 (0.17)
<i>Booralana tricarinata</i>	Small isopod	8	656	12.52 (0.92)	-15.68 (1.96)	3.16 (0.09)
<i>Heterocarpus ensifer</i>	Armed Nylon Shrimp	8	600 - 774	8.30 (0.71)	-14.23 (1.68)	2.85 (0.08)

**Fig 1.** Isotope bi-plot highlighting mean  $\delta^{13}\text{C}$  and  $\delta^{15}\text{N}$  ( $\pm$  SD) from the Exuma Sound fish (black squares) and invertebrate (open circles) community.

**Fig 2.** Deep-water species sampled from the Exuma Sound island slope community: a) *Squalus cubensis* (photo by: Brendan Talwar); b) *Heterocarpus ensifer*; c) *Centrophorus* spp.; d) *Bathynomus giganteus*.

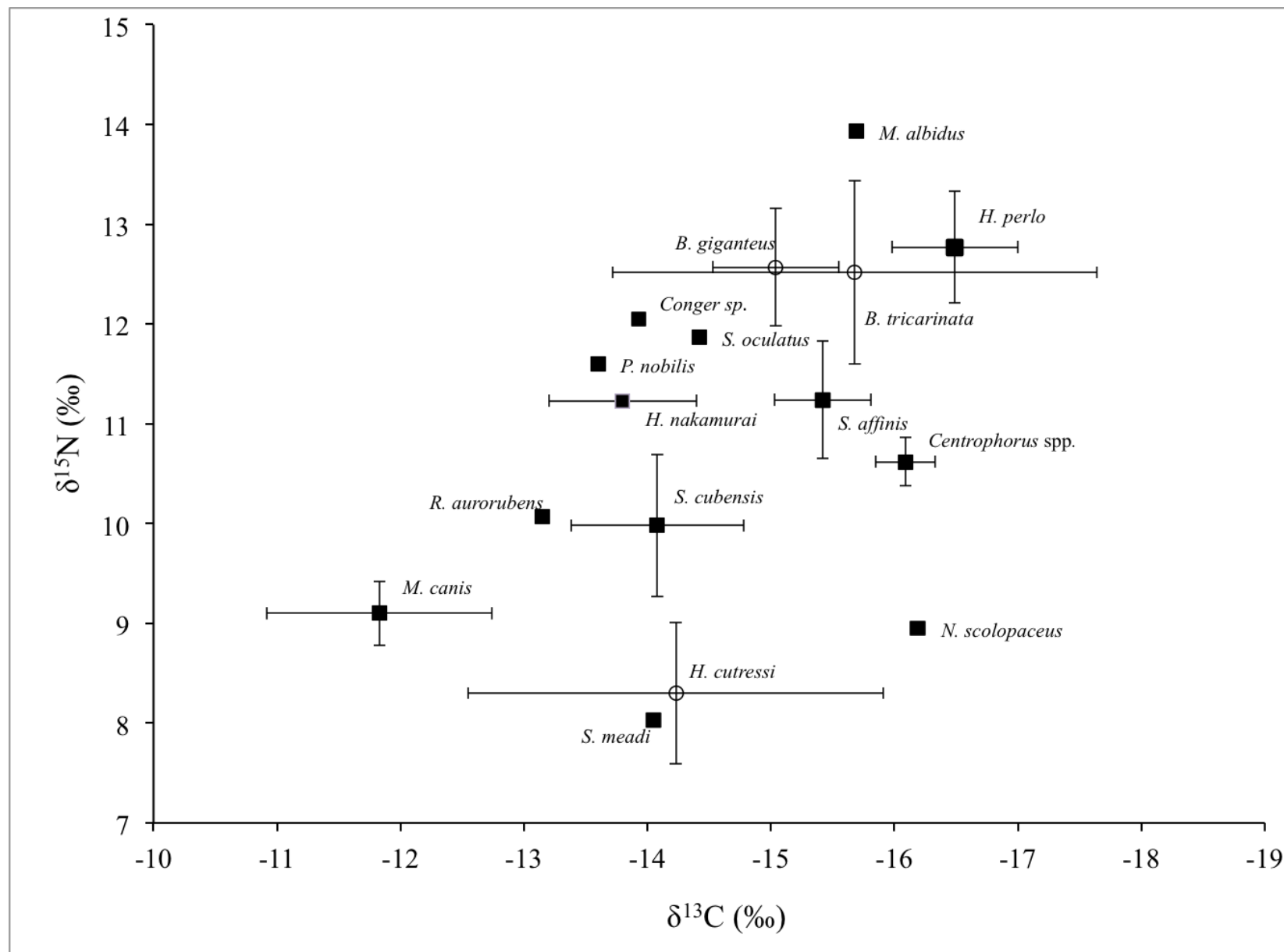


Figure 1

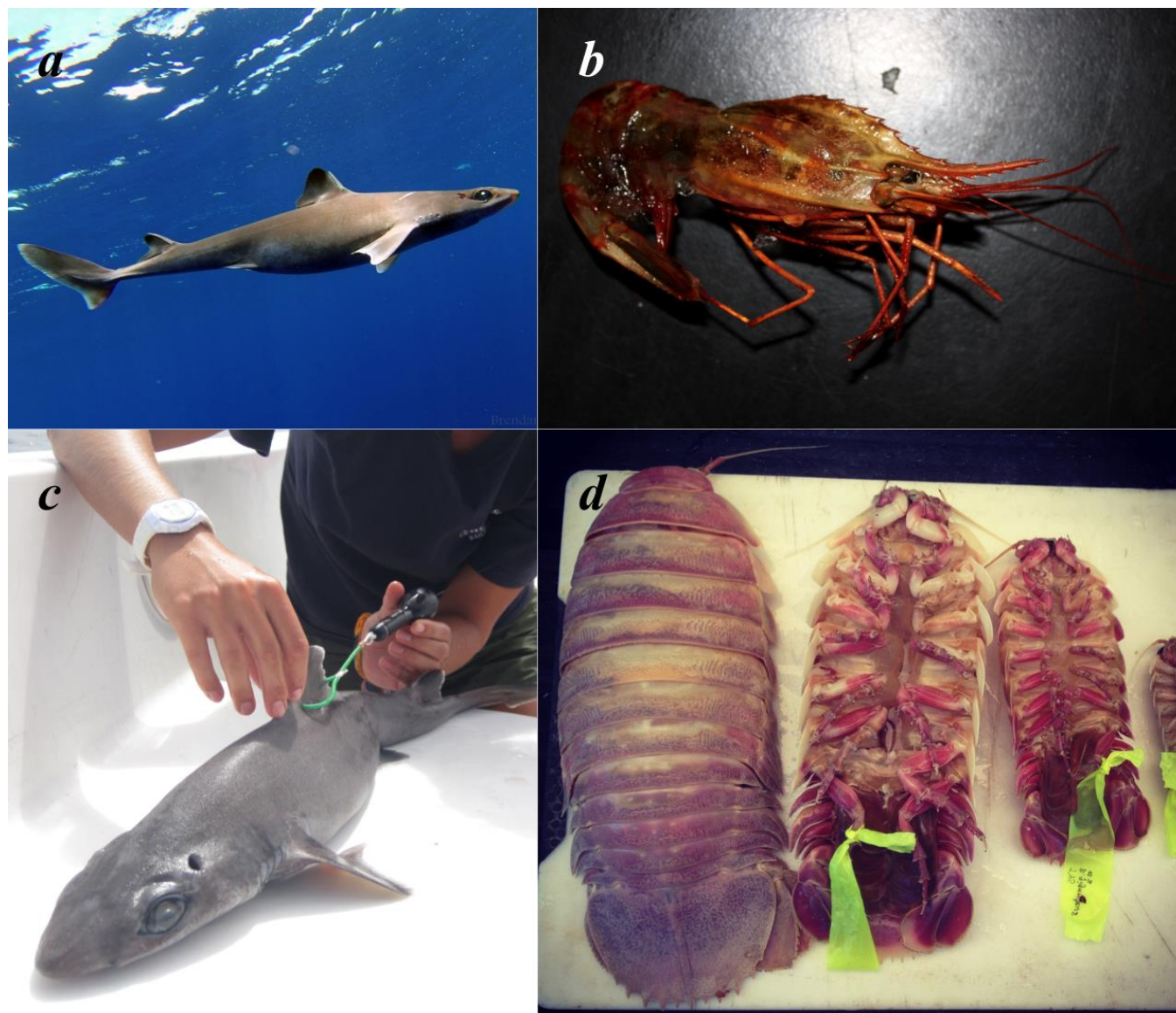


Figure 2